

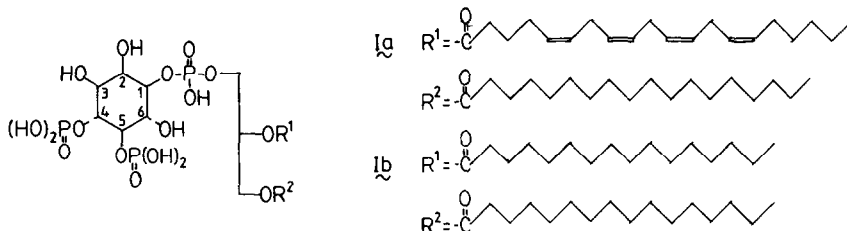
SYNTHESIS OF 1-O-(1,2-DI-O-PALMITOYL-SN-GLYCERO-3-PHOSPHO)-D-MYO-INOSITOL 4,5-BISPHOSPHATE:
 AN ANALOGUE OF NATURALLY OCCURRING (Ptd)Ins(4,5)P₂

C.E. Dreef, C.J.J. Elie, P. Hoogerhout, G.A. van der Marel and J.H. van Boom*
 Gorlaeus Laboratories, P.O. Box 9502, 2300 RA Leiden, The Netherlands

Abstract: Optically active 2,3,6-tri-O-benzyl-4,5-di-O-(*trans*-prop-1-enyl)-D-*myo*-inositol and 1,2-di-O-palmitoyl-*sn*-glycerol were coupled using mono- and bifunctional phosphitylating reagents to yield, after final removal of all benzyl-protecting groups the chiral title compound.

The naturally occurring triphosphoinositide Ia¹ [(Ptd)Ins(4,5)P₂] is believed to play a pivotal role in receptor-mediated Ca²⁺ mobilization. Receptor activation coupled with the action of a specific phospholipase C leads to phosphodiesteratic cleavage of phosphatidyl inositol 4,5-bisphosphate (Ia) into the corresponding diglyceride (1,2-DG) and *myo*-inositol 1,4,5-trisphosphate [Ins(1,4,5)P₃]. The two species thus released by receptor-initiated cleavage may function as second messengers. Thus 1,2-DG was found² to be involved in the activation of protein kinase C, and Ins(1,4,5)P₃ in binding to a receptor, presumably a component of the endoplasmic reticulum, resulting in the discharge of Ca²⁺ from intracellular stores into the cytosol³.

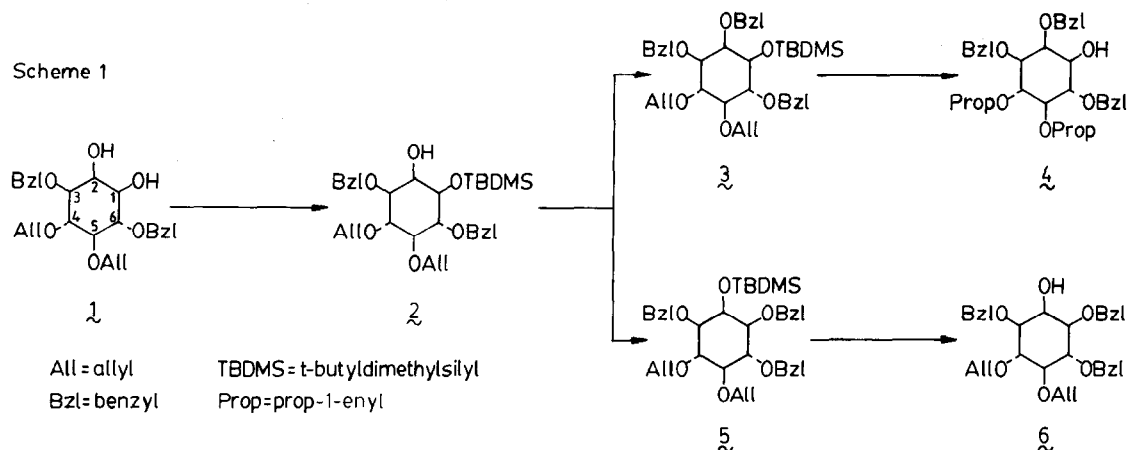
In order to get a deeper insight into the biological function and biosynthetic pathways of (Ptd)Ins(4,5)P₂, we report here a convenient route to the preparation of an optically active analogue of (Ptd)Ins(4,5)P₂ (i.e. compound Ib).



Diastereoisomers of Ib were prepared earlier⁴ in a reasonable yield by a non-regioselective phosphorylation of racemic 3,6-di-O-benzyl-4,5-bis-O-(dianilinophosphoryl)-*myo*-inositol with racemic 1,2-di-O-stearoyl-glycero-3-phosphate in the presence of 2,4,6-triisopropylbenzenesulfonyl chloride, and subsequent removal of all protective groups.

In our approach (see Scheme 1⁵) we started from optically active 4,5-di-O-allyl-3,6-di-O-benzyl-D-*myo*-inositol (1), which was prepared according to the procedure of Ozaki *et al.*⁶. Treatment of 1 with excess *t*-butyldimethylsilyl chloride in pyridine for 16 h at 50°C gave, after purification by short-column chromatography, the crystalline silyl derivative 2 $[\alpha]_D^{20}$ -7.7°, *c* 1, CHCl₃; m.p. 58.5-60°C (from pentane) in a yield of 87%. Benzylation of 2 (10 mmol) in DMF (50 ml) with benzyl bromide (11.5 mmol) and sodium hydride (15 mmol) showed, after work-up, the presence (TLC-analysis) of the two positional isomers 3 and 5 (ratio 4:1), the separation of which could easily be effected by short-column chromatography. The

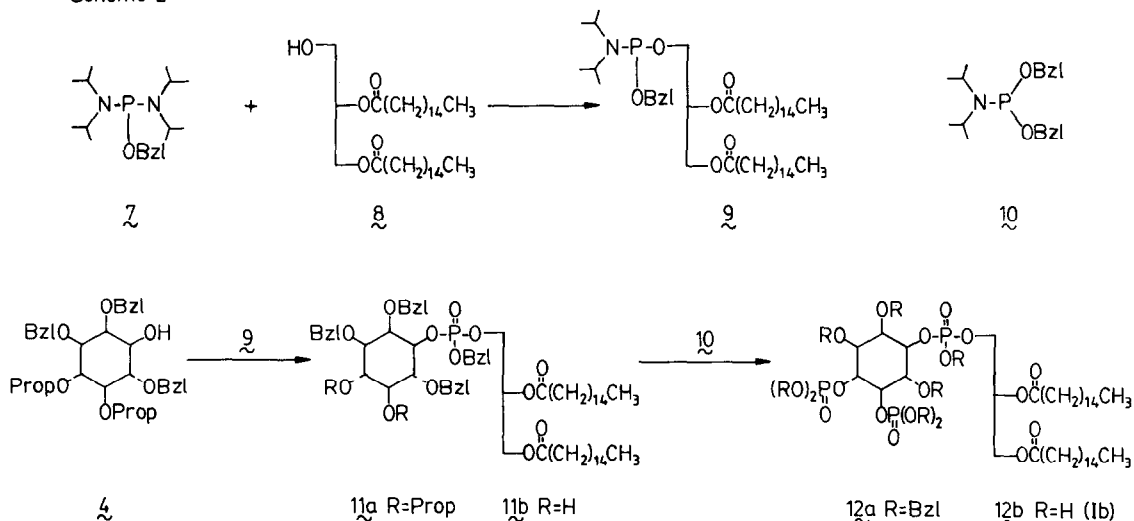
Scheme 1



structural assignment of the minor product 5 ($[\alpha]_D^{20}$ -4.7° , c 1, CHCl_3 ; m.p. $35.5\text{--}36^\circ\text{C}$; 19% yield) was, apart from spectroscopic data (^1H - and ^{13}C -NMR), corroborated as follows. Complete removal of the *t*-butyldimethylsilyl group from the axially orientated hydroxyl group at C-2 in 5 to provide 6 ($[\alpha]_D^{20}$ $+0.3^\circ$, c 1, CHCl_3 ; m.p. $73\text{--}74^\circ\text{C}$) proceeded most conveniently with tetra-*n*-butylammonium fluoride⁷/dioxane at elevated temperature (*i.e.*, 1 h at 50°C). On the other hand, deblocking of the silyl group from the equatorial hydroxyl in 3 with *n*- Bu_4NF proceeded, as expected, rapidly at 20°C . Further, regioselective benzylation of the 1,2-*O*-stannylated⁸ derivative of starting compound 1 with benzyl bromide in the presence of *n*- Bu_4NI ⁹ gave an optically active *myo*-inositol derivative (yield 76%) having identical physical properties as 6 obtained by de-silylation of 5. The required compound 3 ($[\alpha]_D^{20}$ $+7.9^\circ$, c 1, CHCl_3), which was isolated in a yield of 78%, was now converted into the key intermediate 4 by the following two-step one-pot procedure. Isomerization¹⁰ of the allyl-groups into the isomeric *trans*-prop-1-enyl groups could be effected smoothly using a catalytic amount of 1,5-cyclooctadiene-*bis*[methylidiphenylphosphine]iridium hexafluorophosphate¹¹ (activated under H_2 for 2 min) in $\text{ClCH}_2\text{CH}_2\text{Cl}$, followed by removal of the *t*-butyldimethylsilyl group by treatment with *n*- Bu_4NF at room temperature. In this way, crystalline 4 ($[\alpha]_D^{20}$ -6.2° , c 1, CHCl_3 ; m.p. $86.5\text{--}88^\circ\text{C}$ (from pentane)) was isolated in a yield of 90%.

The next stage in the synthesis of Ib consisted of the introduction of one phosphodiester and two phosphomonoester functions. The formation of the phosphodiester linkage between 1,2-*O*-diacyl-*sn*-glycerol 8¹² and the *myo*-inositol derivative 4 *via* a phosphite coupling approach¹³ is outlined in Scheme 2⁵. In the first step, treatment of 1,2-di-*O*-palmitoyl-*sn*-glycerol (8, 2.5 mmol) in CH_2Cl_2 (10 ml) with the bifunctional phosphitylating reagent 7¹⁴ (2.75 mmol) in CH_2Cl_2 (5 ml) and in the presence of 1*H*-tetrazole (1.37 mmol in 2.7 ml CH_3CN) for 30 min at 20°C afforded, after work-up and purification by column chromatography¹⁵, homogeneous 9 (δ_{p} 148.96 and 149.14 ppm) in a yield of 92%. Subsequent coupling of 9 (1.2 mmol) with 4 (1 mmol) in the presence of 1*H*-tetrazole (1.32 mmol) gave an intermediate phosphite-triester (δ_{p} 140.64 ppm). The latter was oxidised *in situ* with *t*-butyl hydroperoxide¹⁶ for 45 min at 0°C . Work-up of the reaction-mixture gave 11a (δ_{p} -1.24 ppm). Acidic hydrolysis (0.1 N HCl/MeOH ; 15 min at 20°C) of the *trans*-prop-1-enyl groups in 11a afforded compound 11b (δ_{p} -1.15 and -1.24 ppm) in an overall yield of 72% based on 4. 1*H*-

Scheme 2



Tetrazole-mediated phosphitylation (see Scheme 2⁵) of 11b (0.5 mmol) with the monofunctional phosphitylating reagent 10¹⁷ (1.5 mmol) afforded an intermediate containing two phosphite-triesters and one phosphate-triester (δ_p 142.55, 141.58 and -1.00 ppm). Oxidation of the latter two phosphite-triester intermediates, as mentioned before, gave after purification by short column chromatography the fully benzyl-protected derivative 12a (δ_p -1.15, -1.27 and -1.36 ppm) in a yield of 83%. Finally, hydrogenolysis ($\text{H}_2/\text{Pd}(\text{C})/\text{CHCl}_3\text{-MeOH}$) of compound 12a for 24 h under pressure furnished 12b (δ_p 0.59, 0.26 and 0.09 ppm), which was isolated as the cyclohexylammonium-salt⁴ in an excellent yield.

In conclusion, the approach described in this paper promises to be a convenient protocol for the synthesis of biologically important analogues of (Ptd)Ins(4,5)P₂. In this respect it is also of interest to note that Ib [$\text{R}^1 = \text{R}^2 = \text{C}(\text{O})(\text{CH}_2)_{14}\text{CH}_3$] could be completely converted with phospholipase A₂ into the corresponding lyso-(Ptd)Ins(4,5)P₂ [*i.e.* Ib $\text{R}^1 = \text{H}$ and $\text{R}^2 = \text{C}(\text{O})(\text{CH}_2)_{14}\text{CH}_3$]. The above lyso-derivative may also give access to analogues of (Ptd)Ins(4,5)P₂ having specific labels (*e.g.* fluorescence or photoaffinity) at the *sn*-2-position of the glyceride moiety. These probes may *inter alia* be useful to study in detail the transport of phosphoinositides between the intracellular membranes by phospholipid transfer proteins 18.

REFERENCES AND NOTES

1. R.C.M. Dawson and J.C. Dittmer, *Biochem. J.*, **81**, 540 (1961). J. Folch, *J. Biol. Chem.*, **146**, 35 (1942). R.V. Tomlinson and C.E. Ballou, *J. Biol. Chem.*, **236**, 1902 (1961).
2. Y. Nishizuka, *Nature*, **308**, 693 (1984).
3. H. Streb, R.F. Irvine, M.J. Berridge and I. Schulz, *Nature*, **306**, 67 (1983). M.J. Berridge and R.F. Irvine, *Nature*, **312**, 315 (1984).
4. V.N. Krylova, N.P. Gornaeva, V.I. Shvets and R.P. Evstigneeva, *Dokl. Akad. Nauk SSSR*, **246**, 339 (1979). V.N. Krylova, A.I. Lyutik, N.P. Gornaeva and V.I. Shvets, *Zh. Obshch. Khim.*, **51**, 210 (1981).

5. All new compounds were characterized by ^1H - and ^{13}C -NMR, as well as combustion analysis.
6. S. Ozaki, Y. Watanabe, T. Ogasawara, Y. Kondo, N. Shiotani, H. Nishii and T. Matsuki, *Tetrahedron Lett.*, 27, 3157 (1986).
7. R.J. Corey and A. Venkateswarlu, *J. Am. Chem. Soc.*, 94, 6190 (1972).
8. J. Alais and A. Veyrières, *J. Chem. Soc. Perkin Trans. I*, 377 (1981). N. Nagashima and N. Ohno, *Chem. Lett.*, 141 (1987).
9. S.J. Angyal and M.E. Tate, *J. Chem. Soc. (C)*, 2367 (1969). S.J. deSolms, J.P. Vacca and J.R. Huff, *Tetrahedron Lett.*, 28, 4503 (1987).
10. J.J. Oltvoort, C.A.A. van Boeckel, J.H. de Koning and J.H. van Boom, *Synthesis*, 305 (1981).
11. L.M. Haines and E. Singleton, *J. Chem. Soc. Dalton Trans.*, 1891 (1972).
12. R.J. Howe and T. Malkin, *J. Chem. Soc.*, 2663 (1951).
13. Thus far only a few reports on the synthesis of glyco(glycero)lipids involving trivalent phosphorus intermediates have been published. For instance; E.E. Nifant'ev *et al.* [*Sov. J. Bioorg. Chem. (Engl. Transl.)*, 4, 877 (1978) and Mendeleev *Chem. J. (Engl. Transl.)*, 23, 27 (1978)] used benzyloxybis(*N,N*-diethylamino)phosphine, K.S. Bruzik *et al.* [*J. Org. Chem.*, 51, 2368 (1986)] methoxy(*N,N*-diisopropylamino)chlorophosphine and S.F. Martin *et al.* [*Tetrahedron Lett.*, 29, 3631 (1988)] phenoxy- or methoxydichlorophosphine for the preparation of glyco- and glycerophospholipids, respectively.
14. Reagent 7 was prepared by dropwise addition of benzyl alcohol (10 mmol) and triethylamine (10 mmol) to a cooled (0°C) solution of bis(*N,N*-diisopropylamino)chlorophosphine (10 mmol) in ether. After 30 min at 20°C, cold (0°C) *n*-hexane (30 ml) was added, followed by removal of $\text{Et}_3\text{N}\cdot\text{HCl}$ by filtration. The filtrate was concentrated *in vacuo* and the remaining oil was redissolved in CH_2Cl_2 to give a 1 M stock solution of 7 (δ_{p} 123.94 ppm).
15. T. Dörper and E.-L. Winnacker, *Nucleic Acids Res.*, 11, 2575 (1983).
16. J. Engels and A. Jäger, *Angew. Chem. Suppl.*, 2010 (1982).
17. The usefulness of *N,N*-diisopropyl(diethyl) dibenzyl phosphoramidites for the introduction of phosphate monoesters was reported recently by J.W. Perich *et al.* [*Tetrahedron Lett.*, 28, 101 (1987)]. The successful application of this type of phosphorylating reagents was further demonstrated in the synthesis of phosphopeptides [W. Bannwarth *et al.* *Helv. Chim. Acta*, 70, 175 (1987) and H.B.A. de Bont *et al.* *Recl. Trav. Chim. Pays-Bas*, 106, 641 (1987)] and inositol phosphates [K.-L. Yu *et al.* *Tetrahedron Lett.*, 29, 979 (1988), C.E. Dreef *et al.*, *Recl. Trav. Chim. Pays-Bas*, 107, 395 (1988) and G. Baudin *et al.*, *Helv. Chim. Acta*, 71, 1367 (1988)]
18. P.A. Paridon, T.W.J. Gadella, P.J. Somerharju and K.W.A. Wirtz, *Biochemistry*, 27, 6208 (1988).

ACKNOWLEDGEMENT

This investigation was supported by the Netherlands Organization for Scientific Research (NWO). We wish to thank Mr. F. Lefeber for recording the ^1H - and ^{13}C -NMR spectra and Professor K.W.A. Wirtz (University of Utrecht, The Netherlands) for his advise and stimulating discussions.

(Received in UK 26 September 1988)